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the Development of Breast Cancer

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INTRODUCTION

One of the hallmarks of tumor suppressor genes is that both copies must be inactivated for the development of the malignant phenotype. A phenomenon associated with this inactivation is that a single recessive mutation is exposed following loss of a chromosomal region containing the normal copy of genes. This event is usually manifested in tumors as loss of heterozygosity (LOH) when compared to normal tissue from the same individual. Genome screening for LOH in breast cancer has identified chromosome region 1p31 as one commonly (50%) involved in LOH indicating the site of a tumor suppressor gene. By comparison of LOH in a large number of tumors, the minimal region has been defined by two markers which are 500-800 Kb apart. An extended region of loss spans approximately 2 Mbp. The overall aim of this project is to characterize the region of chromosome 1p31 which shows consistent loss of heterozygosity in breast tumors by constructing a BAC contig across the region, searching for genes in this contig and then performing a mutation analysis to determine which ones are involved in tumorigenesis. One gene, TTC4, was identified in this region and, after extensive analysis, could not be shown to be involved in breast cancer. It has since been shown that this gene is mutated and so potentially involved in the development of some melanomas. 1p31 is also frequently involved in LOH in melanomas. To identify more genes in the critical 1p31 region involved in breast cancer, we extended our search for candidates using a variety of approaches but, in particular, using the latest releases of sequence form the Human Genome Project.

BODY

In our previous report we described the construction of a YAC contig across the minimally deleted region which extended approximately 1500 Kb. In the early part of this year we began a series of hybrid cDNA capture experiments to isolate genes from a breast cancer cDNA library. This approach depends on hybridizing cDNA clones from a total library to YAC DNA and then, through successive rounds of amplification and selection, a number of clones are recovered for testing which apparently have homology to the target YAC clone. Because of the presence of repetitive sequences in many cDNA clones, however, the risk of isolating false positives is very high using this approach, especially with YACs since they are, on average, 1 Mbp long. After a series of experiments, we isolated a number of candidate cDNAs but were unfortunately not able to map any of the clones back to the YAC contig used in the screen. At this point, data from the Whitehead Institute describing new releases of the human gene map for chromosome 1, although not identifying any fulllength genes in the critical 1p31 region, did map the progesterone receptor gene (PTGFR) to the distal border of this region. Because of the potential involvement of this receptor in breast cancer development, we decided to submit this gene to an analysis by SSCP/mutation screening and DNA sequencing. In order to establish the exon/intron structure, we identified a BAC clone which contained the PTGFR gene and then, by aligning the cDNA sequence with the genomic sequence, were easily able to establish the exon structure. The PR gene only contains three exons and so primers were designed (Table 1) to amplify the individual exons and subject them to SSCP analysis. This approach identifies single strand PCR products which show altered mobility on acrylamide gels, indicative of a DNA sequence change (either mutation or polymorphism). Since each of the PTGFR exons were large (>250 bp), we designed a series of overlapping PCR products to cover each exon (Table 1). We used 32 tumor samples from breast cancers in this analysis and identified a number of band changes. When sequenced, these changes proved to be silent polymorphic variants resulting

in silent changes in the amino acid code for the gene (Table 2). From this survey, therefore, we concluded that PR gene is not frequently mutated in breast cancer.

At this point the accelerated pace of the human genome sequencing project dictated an alternative approach, which was to build up BAC contigs from sequence databases across the 1p31 region and then use database searching to identify ESTs and genes. Using the available physical map which was used to created the YAC contig, we searched the high-throughput genome sequencing database for BACs across the region. These are shown in Figure 2. These individual BACs were identified and then overlapping BACs from the same databases were identified by BLAST searches. The sequencing effort is being undertaken by the Sanger Center in the UK and the genome center at Washington University. Since there is considerable redundancy in these two efforts, it was possible to build up a contig of BACs with only four gaps, the lengths of which cannot be determined at this stage. Of the 2-3 Mbp region, approximately 50% has been sequenced; thus, there is adequate sequence to begin to build up a transcription map of the region to search for candidate genes.

Using BLAST searches and the Genome Channel program (GenScan) at Oak Ridge National Laboratories, we were able to identify ESTs from the region. These are summarized in Table 3. Firstly, It has been an important observation that there are few genes for which the full-length sequence to map this region is known. A number of ESTs were identified, but most of these cluster at the proximal end of the contig and include the latrophilin gene which has previously been shown not to be mutated in breast cancer cells. The only other gene in this region is the DRIL2P gene which is a pseudogene for the DRIL transcription factor. The distal 60% of this contig apparently contains no ESTs, either through BLAST searches or through analysis of the sequence from the individual BACs using GenScan. Although a number of individual exons are predicted in this region using GRAIL, no extensive gene structures could be identified. Even though these gene prediction tools are notoriously inaccurate in their ability to identify genes, it is still true that the region is very gene poor which has the advantage that any gene which is found has a higher probability of being the one reduced to homozygosity in breast cancers.

Within the proximal clusters of ESTs, a number show homologies to Unigene clusters which provide for more cDNA sequence from these genes. Despite this, the available sequence still does not provide any indication of the potential function of any of these genes. We are now concentrating on using the available sequence information to screen cDNA libraries to identify more full-length clones.

KEY RESEARCH ACCOMPLISHMENTS

- Construction of a BAC contig across the LOH region in 1p31.
- Analysis of approximately 1700 kb of sequence using GenScan to predict genes in the critical region.
- Identification of ESTs and Unigene clusters in the 1p31 region
- Exclusion of the progesterone receptor gene in the development of breast cancer

REPORTABLE OUTCOMES

None

CONCLUSIONS

As a result of advances in the development of the human genome sequencing project, we have abandoned traditional gene cloning strategies and have concentrated on building up the contig of complete sequence data for the critical BRCA region in 1p31. This analysis

identifies approximately 1.7 Mbp of the region with only 4 gaps. An analysis of this data places the progesterone receptor at the outer limit of the contig and mutation analysis of this gene does not support its role in the consequences of LOH. We have identified 27 ESTs to date in the critical region and five Unigene clusters. None of the data available for genes in this region allows for a prediction of their function. The construction of a rudimentary transcription map for the 1p31 region involved in breast cancer now means that the isolation of full length genes will be possible by cDNA library screening. As the sequence data become more complete, the generation of a full transcription map will become possible.

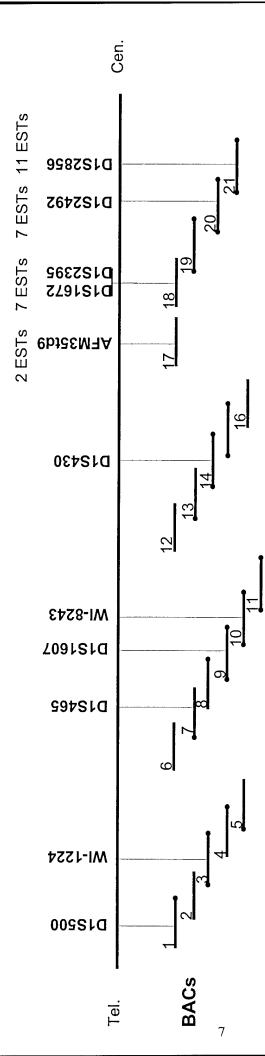
REFERENCES

None

APPENDICES

None

Figure 1: Summary of the BAC map of the BRCA 1p31 region relative to the physical map. The location of ESTs identified by BLAST searches are shown.



1: RP11-239A21, Acc# AC025995

2: RP11-456F13, Acc# AL358533 3: RP11-316O2, Acc# AC073086

Acc# AL360080 RP11-466L22, Acc# AL355876 4: RP11-48318, 5: RP11-466L2

Acc# AC024634 Acc# AL356501 RP5-1159G19, RP5-887A10, RP11-3B10, .: .: 8: .:

Acc# AL391062

Acc# AL136234 Acc# AC009936 RP11-115A15, RP5-933B4, . 6

11: RP11-339D23, Acc# AC024634

Acc# AL354949 12: RP11-82L20,

13: RP11-316C12, Acc# AC068047 Acc# AL356856 14: RP1-54E18.

Acc# AC027532 15: RP11-115M14, Acc# AC073627 16: RP11-417C2,

17: RP3-445O10, Acc# AL158173

18: RP5-837I24, Acc# AL138799

20: RP11-100D1, Acc# AL358939 19: RP4-653E17, Acc# AL138841

21: RP11-211B4, Acc# AL359705

Table1: Primers sequences, PCR product size and PCR conditions used for SSCP analysis for PTGFR gene.

Primer name	Sequence	PCR product Size	Annealing T°C
PTGFREX1F PTGFREX1R	AAC GAG TGC GCG GAG GGG AC CTG GTC CAA ACT CAG TGT CAG G	300bp	65
PTGFREX2-1F PTGFREX2-1R	GGC CAG ATA AAA CCC ATC CC AAG ATT CCC ACT GTC ATG AAG A	302bp	57
PTGFREX2-2F PTGFREX2-2R	CCT GCA GCT GCG CTT CTT TC CCA TGC AGA TAC CAA AAA TAC TG	313bp	09
PTGFREX2-31F PTGFREX2-3R	CTC ATC AAT GGA GCC ATA GC CGC CTG AAT TTT ATA GTC TCG	300bp	53
PTGFREX2-4F PTGFREX2-4R	CGA AAA TTA CAT CCA AAC AT GAG CTG GAT TAC CAT TTC CA	320bp	53
PTGFREX2-5F PTGFREX2-5R	ATG CAA TCA CAG GAA TTA CAC CTG AGT GCT GAA ATA AAT TTT G	265bp	55
PTGFREX3-1F PTGFREX3-1R	AAA CAA TAG CAT CAC TCT GTG CAT GCA CTC CAC AGC ATT GA	300bp	09
PTGFREX3-2F PTGFREX3-2R	CCGAAT GGC AAC ATG GAA TC AGG TAT TTA ACT AAC TGA AAT ATTG	290bp	55

Exon-Intron Boundries for PTGFR gene were determined by aligning the cDNA sequence of PTGFR with the unfinished sequence of BAC clone RP5-944H6 (Acc# AL136324).

Table 2: Summary of the silent polymorphisms found in the PTGFR gene in breast cancers. The 2 polymorphisms with a minus (-) annotation occur within the intron for exon 2.

Exon	nucleotide	amino acids
2	C63T	T21T
2	-152 T ->	C
2	−62 T ->	C
2	G435A	T145T

BAC clone Acc#	EST Acc#	UniGene Cluster	Gene name
RP3-445O10 AL158173	AA909664 AA827584	Hs.132428	
RP5-837I24 AL138799	R10940 AI147471 AA406063 BE894577 BE279803, BE256076 AI267423	Hs.146741 Hs.98003	DRIL2P
RP4-653E17 AL138841	AI732798, AA487326 AW105209	Hs.188839 Hs.242795	
RP11-100D1 AL358939	AW270271, AI243907 AA347902 W03266 AA347901 N69441	Hs.149086	
RP11-211B4 AL359705	AI005162 AA369954 R01807 AA347901 AA259171 BE066265 T06662 H63565 H63611 BE709816		
	AL040055	Hs.24212	LATROPHILIN

Table 3: List of EST markers and their respective UniGene clusters that map to BAC clones from the breast cancer critical region in 1p32.